

석사학위논문

**Niemann Pick Disease , type C 생쥐 해마로
투사하는 중추신경로의 변화에 관한 연구**



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ABSTRACT

Retrograde transneuronal labelling of the pseudorabies virus was used to investigate the differences of the CNS hippocampal pathways among NPC +/+, +/-, and -/- mice. For hippocampal pathways, a total of 3-5 ul Bartha strain of pseudorabies virus were injected into the hippocampus using stereotaxic instrument in 40 adult NPC +/+, +/-, and -/- mice. After 48 hours post pseudorabies virus injection, the brains were removed and the tissue sections were processed for immunohistochemistry using polyclonal antibody against the pseudorabies virus. Using stereological counting methods, the PRV positive cells were counted in the positive nuclei of the whole brain of the NPC +/+, +/-, and -/- mice. The relative number of PRV positive cells in labelled nuclei were same in the hippocampus and medial septal nuclei of the NPC +/+, +/-, -/-. But there were significant decrease of PRV positive cell number in entorhinal cortex, cerebral cortex including piriform

cortex, lateral septal nucleus, thalamus, lateral hypothalamic nucleus, nucleus of the diagonal band, superior mammillary nucleus, locus cereuleus, medial geniculate nucleus, anterior olfactory nucleus in NPC $-/-$ compared to the $+/+$ and $+/-$, and some showed no positive labeling. In summary, the CNS hippocampal pathways of the NPC $-/-$ were reduced and the number of the cells within the pathways decreased in a circuit specific manner.

Our data suggests that the discontinuity of CNS hippocampal pathways in NPC $-/-$ mice may contribute to the symptoms and functional disabilities especially memory and learning.

Keywords : Niemann-Pick type C, hippocampus, CNS pathway, pseudorabies virus

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INTRODUCTION

Niemann-Pick type C disease (NP-C) is a fatal, autosomal recessive disorder that results in premature death from progressive neuro-degeneration. (Penchev et al., 1995) Recently, researchers have discovered two genes that can produce the NP-C phenotype when mutated. In 1997, NPC1 was identified and sequenced (Carstea et al., 1997; Loftus et al., 1997) and in 2000, a second gene (HE1/NPC2) was identified (Naureckiene et al., 2000). At the cellular level, mutations in the NPC1 and NPC2 genes result in the late lysosomal accumulation of lipids [Sokol et al., 1988; Blanchette et al., 1988] and ultimately lead to disorders of cholesterol imbalance. NPC patients suffer from such symptoms as ataxia, impaired vertical gaze, and dementia. The common neuropathological features of NPC include neuronal ballooning, axonal spheroid formation, and neuro-degeneration (Tanaka et al., 1988; Penchev et al, 1995, Higashi et al., 1991, Higashi et al., 1993). The extent and specificity of neurodegeneration is still largely unknown.

The hippocampus is widely believed to exert an important influence on memory. And several neuronal pathways including the septohippocampal system are thought to play a crucial role in the

performance of a variety of complex learning and memory tasks[Coyle et al., 1983; Lamour et al., 1984]. This specific hippocampal function can be performed by neuronal circuits which connected in the brain. In cognitive and memory processes, cholinergic involvement has been known as one of the most important transmitter system.

The BALB/c-npc1^{nih} mouse model for Niemann-Pick type C has facilitated closer examination of this phenomenon of abnormal cholesterol storage and neuronal loss in the central nervous system. As in the human phenotype, central neurological symptoms such as ataxia and hind limb paresis dominate the clinical picture.

In order to assess the neuro-degenerative effects of NP-C on hippocampal afferent pathways, we performed a pseudorabies virus neurotracing study on wild-type NPC1 mice (+/+) and NPC1 knockout mice (+-)/(-/-).

Materials and Method

1. Animals The BALB/c mice adult wild-type control(NPC +/+), heterozygous(NPC +/-) and homozygous(NPC-/-) animals of both sexes were used in this study. We have established a NPC mice colony, BALB/c mice carrying the genetic mutation for NPC1, and are ready to be used for our experiments. NPC heterozygous mice are being bred to acquire NPC (+/+), NPC (+/-), and NPC (-/-) mice. The genotypes of the mice are determined from genomic DNA isolated from tail-snip DNA using a polymerase chain reaction (PCR)-based method and oligonucleotide primers described previously.

2. Pseudorabies virus The Bartha strain of the pseudorabies virus(PRV) was used in this study. The titer of the virus stock was, determined on PK15 cell line, was 1×10^8 plaque forming unit(pfu)/ml. Aliquots of the PRV were stored at -80 oC, and vials were thawed immediately prior to the injection. Excess virus were inactivated with Clorox and discarded.

3. Surgery Animals were anesthetized with ketamine HCl(0,75 mg/kg) and xylazine(1 mg/kg) prior to surgical procedures The injection of pseudorabies virus was made into the hippocampus with an aid of the stereotaxic instrument at the point from bregma [dentate gyrus , 3.7

mm(AP), 2.4 mm(Lateral)]and Hamilton syringe was lowered vertically until it reaches the injection areas. Total 3 ul of the pseudorabies virus was injected slowly at the speed of 1 ul per minute. Then syringe was removed slowly and surgical wounds were sutured with wound clips. Most of the rats were allowed to survive a total of 2 days postinjection.

4. Immunohistochemistry All mice were reanesthetized in same manner and perfused transcardially with 100-200 ml of heparinized saline(18 oC) followed by 400 ml of 4% paraformaldehyde-lysine periodate. The brains were removed, and cut in transverse plane at 30 um on a freezing microtome. The free floating tissue sections were processed for double immunofluorescence procedure using polyclonal antibodies to pseudorabies virus or choline acetyl transferase. One every six sections was incubated overnight at 38 oC with the mixture of rabbit anti pseudorabies virus(1:200) and goat anti choline acetyl transferase(1:200, Chemicon Int'l Inc., diluted in 0.1 M sodium phosphate buffer containing 1% normal donkey serum and 0.3% Triton X-100). After 14 hours, the sections were reacted for 2 hours with a cocktail of FITC labelled donkey anti-rabbit IgG(1:50, Jackson Immunoresearch Lab.) and TRITC labelled donkey anti-goat IgG(1:50, Jackson Immunoresearch Lab.). Cells were counted as positive if a reasonable portion of the cell body was visible in the section.

5. Stereology CAST stereology system with Olympus BX-51 microscope and CAST software were used for stereological cell counting of the PRV positive neurons in all mice. The unbiased stereological estimation of the total number, cell size and axon populations were made the optical fractionator. All the nuclei containing positive neurons were outlined from front to back through the nucleus/region of positive labelled. The PRV positive labelled cells were counted in regions outlined. Sampling was done using the Olympus CAST(Computer Assisted Stereological Toolbox) system version 2.1.0(Olympus Denmark A/S, Albertslund, Denmark). A counting frame($2025 \text{ } \mu\text{m}^2$) was placed randomly on the first counting area and systematically moved through all counting areas until the entire delineated area was sampled. The sampling frequency was chosen so that about 140 PRV positive cells were counted in each specimen. Average thickness of the sections were 12-17 μm . A neuron was defined as a cell with a clearly visible cell body within the 10 μm Z-plane of the counting frame using a 100X oil immersion objective. An upper guard zone of 2 μm was used, so the cells were excluded from both surfaces to avoid the problem of the lost caps, and only the profiles that came into focus within the counting volume were counted. The estimate of the total number of the PRV containing neurons were calculated according to the optical fractionator formula.

6. Statistics Analysis of variance(ANOVA) was used compare the

percentage cell number (mean + S.E.M.) of the NPC^{+/+}, ^{+/-}, and ^{-/-} animals.
The P<0.05 level was used determine statistical significance.



RESULT

1. Cresyl violet staining in the NPC +/+, +/- and -/- mouse hippocampus

As shown in Table 1, at 4 weeks and 8 weeks of age both NPC -/- and NPC +/+ mice showed no significant differences in stereological cell counts of cresyl stained cells in the CA1, CA2/3 and dentate gyrus regions of the hippocampus.

2. PRV positive cells in the NPC +/+, +/- and -/- mouse hippocampus

After injection of pseudorabies virus (PRV) into the hippocampus, the PRV immuno-positive cells were observed in several different areas of the brain. The labeling pattern of NPC -/- was different to NPC +/+ and +/- . The relative number of PRV positive cells in labelled nuclei were increased in the hippocampus(Hp), (HDV), and medial septal nucleus(MS) of the NPC-/- mouse.

But there were significant decrease of PRV positive cell number in entorhinal cortex(E), Cerebral cortex(CC) including

piriform cortex(P), lateral septal nucleus(LSD), thalamus(TH), lateral hypothalamic nucleus(LH), nucleus of the diagonal band(VDB), locus ceruleus(LC), medial geniculate nucleus(MG), dorsal entorhinal nucleus(DEn) in NPC $-/-$ compared to the $+/+$ and $+/-$, and some showed no positive labeling.



DISCUSSION

The Pseudorabies virus (PRV) is a swine neurotropic alpha-herpesvirus that has been used for over twenty years to map neural circuits due to its ability to cross synaptic junctions. (Loewy et al., 1998) The Bartha strain, an attenuated strain used for retrograde tracing, can be injected into the peripheral nervous system or intracerebrally (Card J.P. et al., 1999; Aston-Jones G. et al., 2000). PRV has been used to map such pathways as: the retinal projections in mice (Provencio I. et al., 1998), the rat nucleus accumbens and thalamus (O'Donnell P. et al., 1997), and the rat prefrontal cortex to the ventral tegmental area (Carr D. B. et al., 2000). But care must be taken when using this virus to label neural circuits that are undergoing neuro-degeneration as is the case with NPC.

Research on PRV-Bartha's replication, assembly and egress in neurons (Card et al., 1993) can shed light on its uses and limitations as a neurotracer in neurodegenerative cases. Viral replication occurs within the nucleus of the infected cell. From here, the naked capsid buds from the nuclear envelope and transverses the endoplasmic reticulum (ER). As the nuclear envelope fuses with the ER membrane the naked capsid is freed into the cytoplasm near

the Golgi complex. The Golgi complex then wraps around the naked capsid to form a bilaminar envelope. The enveloped capsids then enter the dendritic portion of the neurons where the outer membrane of the envelope can fuse with the post-synaptic terminal. The capsid, now surrounded by a single membrane, crosses the synaptic junction and fuses to the pre-synaptic terminal. The naked capsid then travels up the axon towards the nucleus to repeat the process of replication and egress. This process of infection must be kept in mind when using it as a neurotracer in NPC tissue

There are several possible explanations for the loss of PRV labeling in NPC1 (-/-) mice. (1) NPC1 (-/-) mice accumulate a large number of cholesterol-loaded vesicles within the cell body that can hinder viral movement throughout the neuron. This traffic jam effect could have an affect on viral access to the nucleus and/or on viral egress from the cell. (2) Intracellular trafficking of cholesterol is hindered in NPC (-/-) mice. If Golgi derived enveloping of viral capsids is cholesterol dependant then this process would be hindered in NPC1 (-/-) mice. (3) If neurodegeneration has reached a point where synapses are lost then the virus would be unable to enter the crippled neuron. (4) If non-labeled neurons have undergone nuclear lysis this would hinder viral entry, which appears to be receptor mediated, and would hinder viral replication.

In the present study, we examined the transsynaptic cholinergic circuitry projecting to the hippocampus using the pseudorabies virus as a neurotracer. Decreased nuclei are entorhinal cortex(E), Cerebral cortex(CC) including piriform cortex(P), lateral septal nucleus(LSD), thalamus(TH), lateral hypothalamic nucleus(LH), nucleus of the diagonal band(VDB), locus ceruleus(LC), medial geniculate nucleus(MG), dorsal entorhinal nucleus(DEn). This may be because of the difficulties in the viral transport due to the cholesterol accumulation in the npc ^{-/-} hippocampal cell body. Interestingly, hippocampus(Hp), (HDV), and medial septal nucleus(MS) showed increased immunoreactivity in npc^{-/-} mouse. This may help the explanation that virus had difficulty in the remote transport but increased in the same cells which they infected first. So these nuclei could be candidates of the closely related nuclei to the hippocampus.

Totally, these changes tells that the CNS pathway to the hippocampus has been broken occurs along the pathway, and cause memory disturbance

CONCLUSION

Our data suggests that the CNS pathways innervating the hippocampus were different among NPC +/+, +/-, -/- mice and the hippocampal innervation also showed degenerative changes in a transsynaptic manner in NPC -/- mice. The degeneration, disturbance of the transport in the axon or abnormal function of the synapse could be possible candidate for NPC mice symptoms.



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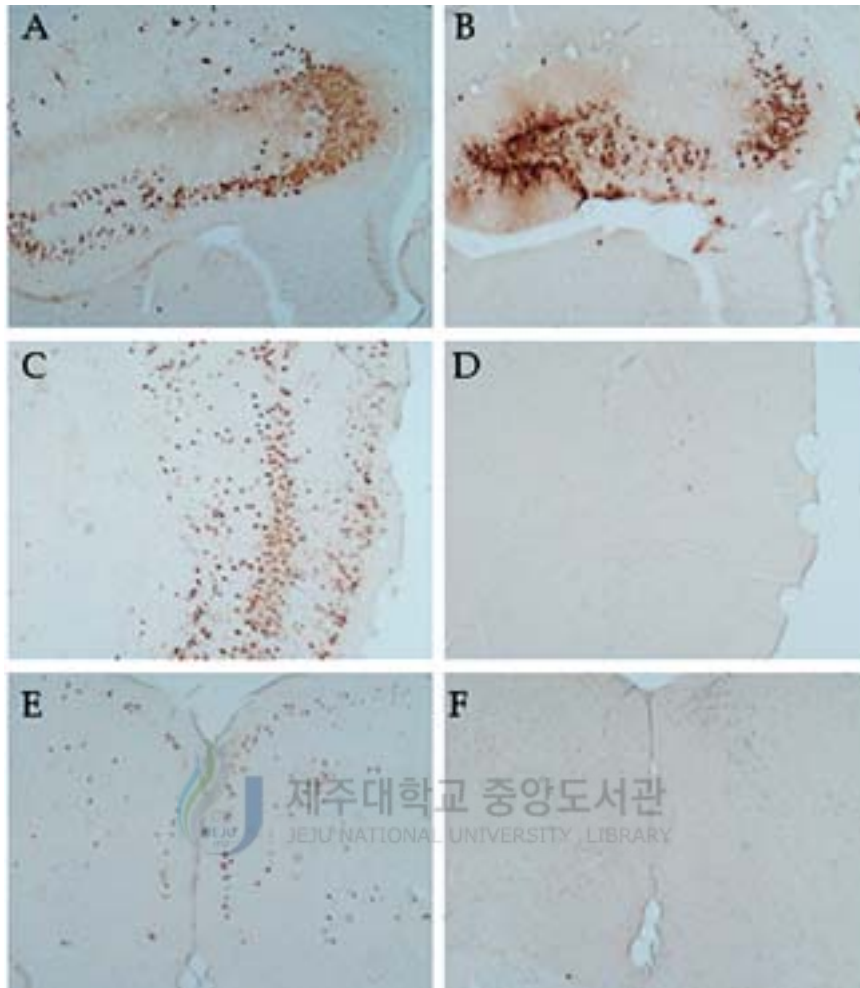
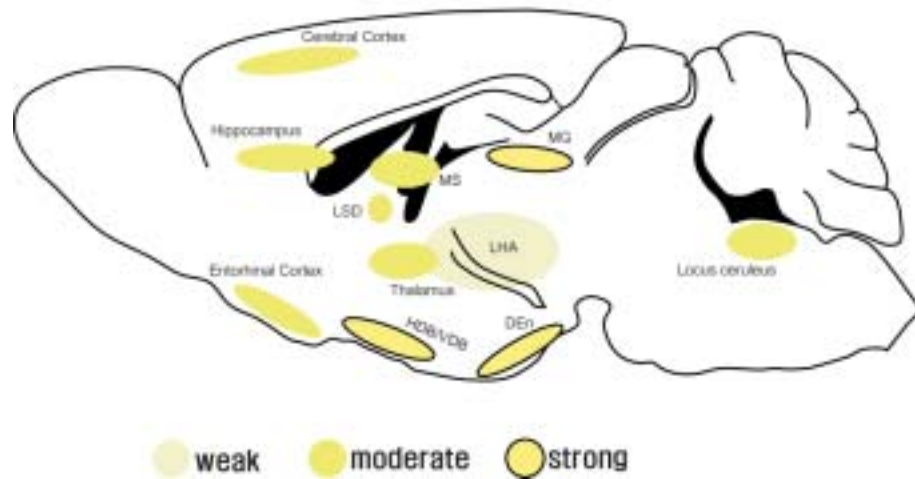


Fig. 1. PRV-immunoreactive neurons in the brain ($\times 40$) a, b: hippocampus; c, d: cerebral cortex; e, f: cingulate cortex. a,c,e : npc +/+, b,d,f : npc-/-

Table 1. The average PRV positive cell number per each section at 48 hrs after PRV injection into the hippocampus of the mouse brain

	+/+		+/-		-/-	
	injection	non injection	injection	non injection	injection	non injection
Hp	30	12	30	16	75	48
E+P	34	8	56	28	11	0.9
C.C	43	29	40	32	1.2	0.7
LSD	32	16	52	20	0	0
TH	26	22	31	12	7	2
LH	17	9	15	8	3	1
HDV	48	25	120	15	59	58
VDB	95	37	88	17	40	16
MS	39	24	54	23	84	61
SUM	-	-	-	-	90	55
LC	39	12	-	-	16	1.6
DEn	76	25	118	56	0	0
MG	53	32	38	20	0	0

Hippocampal Pathway of Mouse after PRV injection



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Fig. 2. Schematic drawings illustrating the positive nuclei after PRV injection into the hippocampus of the NPC +/+ mouse

Alteration of Hippocampal Pathway in NPC $-/-$ Mouse

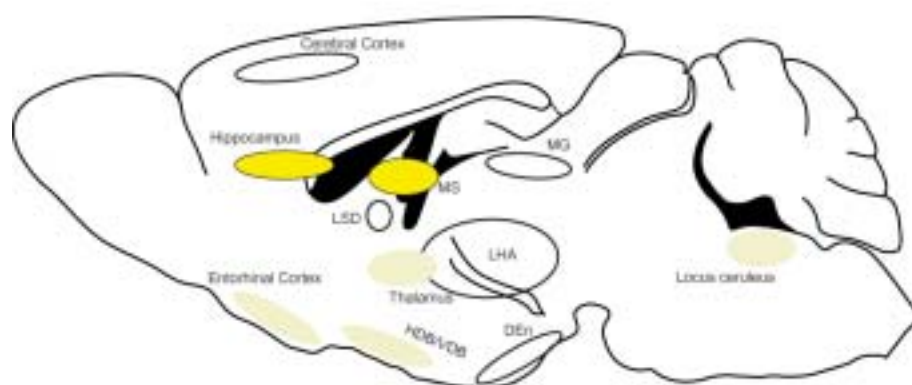


Fig. 3. Schematic drawings illustrating the positive nuclei after PRV injection into the hippocampus of the NPC $-/-$ mouse

Abbreviations : C.C, Cerebral cortex; DEn, Dorsal entorhinal area; E+P, Entorhinal and piriform cortex; HDV, Diagonal band, H; Hp, Hippocampus; LC, Locus ceruleus; LH, Lateral hypothalamic area; LSD, Lateral septal dorsal nucleus; MS, Medial septal nucleus; SUM, supramammillary nucleus; TH, Thalamus; VDB, Diagonal band, Ventral



Niemann Pick Disease , type C

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knock-out pseudorabies virus



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